# KARADENİZ TECHNICAL UNIVERSITY THE GRADUATE SCHOOL OF NATURAL AND APPLIED SCIENCES

# DEPARTMENT OF FISHERIES TECHNOLOGY ENGINEERING

# **GASTRIC EVACUATION OF FORMULATED FEEDS IN BROOK TROUT**

**Ph.D. THESIS** 

**Umar KHAN** 

**JANUARY 2020<br>TRABZON** 



# KARADENİZ TECHNICAL UNIVERSITY THE GRADUATE SCHOOL OF NATURAL AND APPLIED SCIENCES

DEPARTMENT OF FISHERIES TECHNOLOGY ENGINEERING

## **GASTRIC EVACUATION OF FORMULATED FEEDS IN BROOK TROUT**

**Umar KHAN** 

# This thesis is accepted to give the degree of **DOCTOR OF PHILOSOPHY**

By

The Graduate School of Natural and Applied Sciences at **Karadeniz Technical University** 

> The Date of Submission :  $15/12/2019$ **The Date of Examination**  $: 16/01/2020$

Thesis Supervisor : Prof. Dr. Kadir SEYHAN

**Trabzon 2020** 

# KARADENİZ TECHNICAL UNIVERSITY THE GRADUATE SCHOOL OF NATURAL AND APPLIED SCIENCES

# DEPARTMENT OF FISHERIES TECHNOLOGY ENGINEERING **UMAR KHAN**

## **GASTRIC EVACUATION OF FORMULATED FEEDS IN BROOK TROUT**

## Has been accepted as a thesis of

## **DOCTOR OF PHILOSOPHY**

after the Examination by the Jury Assigned by the Administrative Board of the Graduate School of Natural and Applied Sciences with the Decision Number 1833 dated  $24/12/2019$ 

**Approved By** 

- Chairman : Prof. Dr. MİNE KIRKAĞAÇ
- Member : Prof. Dr. KADİR SEYHAN
- Member : Prof. Dr. GÖKTUĞ DALGIC
- Member : Prof. Dr. NADİR BAŞÇINAR

: Assoc. Prof. Dr. RAHŞAN EVREN MAZLUM **Member** 

Prof. Dr. Asim KADIOĞLU **Director of Graduate School** 

## **ACKNOWLEDGEMENTS**

First, I would like to express my immense respect and sincere gratitude to my supervisor, Prof. Dr. Kadir SEYHAN for his indefatigable enthusiasm. I have had the privilege to benefit from his knowledge, professional experience, and wisdom throughout my Ph.D. Dr. Seyhan always showed patience and dedication when helping me with small or big issues, and at the same time gave me the freedom to work independently. I extend my heartfelt gratitude to Prof. Dr. Nadir BAŞÇINAR for his invaluable help which facilitated this research. I also thank Dr. Niels Gerner ANDERSEN from DTU Aqua, Denmark for giving me important statistical advice and helping me in the derivation and application of the general power function and temperature optimum function.

I am also thankful to the "Presidency for Turks Abroad and Related Communities (YTB)" who granted me the opportunity for Ph.D. studies at Karadeniz Technical University, Trabzon. As well, I give special thanks to the Scientific and Technological Research Council of Turkey (TUBITAK; grant no: 113O362) for providing funding which allowed me to undertake this research.

My acknowledgement would be incomplete without thanking my beloved mother for her support and love throughout my life. I also thank my brother Mr. Rahmatullah who looked after our family during the final portions of my study; I appreciate him for the excellent example he continues to set.

And last, but definitely not least, I am greatly indebted to and greatly appreciative for and to the most important person in my life – my wife Zahra. Her patience, understanding, and constant support during this program has been crucial to my success. I dedicate this Ph.D. thesis to both of my lovely daughters, Inaaya, Arya, and to my wife, all of whom are very precious to me and constitute the reasons why I laugh, smile, and want to get up every morning. Also, I wish to offer special admiration to my late hard-working father who literally labored day and night to support the education of his children and who guided us as a responsible, caring, and loving father.

> Umar KHAN Trabzon, 2020

## **STATEMENT OF ETHICS**

I declare that, this PhD thesis, I have submitted with the title "Gastric evacuation of formulated feeds in brook trout" has been completed under the guidance of my PhD supervisor Prof. Dr. Kadir SEYHAN. All the data used in this thesis were obtained experimental works done as parts of this thesis in our research labs. All referred information used in the thesis has been indicated in the text and cited in reference list. I have obeyed all research and ethical rules during my research, and I accept all responsibility if proven otherwise. 05/02/2020

*Amarchan* 

Umar KHAN

# **TABLE OF CONTENTS**





### PhD. Thesis

#### **SUMMARY**

## <span id="page-7-0"></span>GASTRIC EVACUATION OF FORMULATED FEEDS IN BROOK TROUT

Umar KHAN

Karadeniz Technical University The Graduate School of Natural and Applied Sciences Department of Fisheries Technology Engineering Supervisor: Prof. Dr. Kadir SEYHAN 2020, 42 Pages, 12 Pages Appendix

Gastric evacuation rate (GER) can be used to estimate stomach fullness in fish at any postprandial time, to predict return of appetite and optimize feeding regimes. The influences of meal size, fish size *L* (cm), temperature  $T$  (°C), dietary energy density *E* (kJ  $g^{-1}$ ) and feed storage conditions (frozen, ambient conditions), on GER of brook trout fed a single meal of commercial pellets were studied. In addition, the effect of a second meal on GER was examined. Small (15 cm) and large (23 cm) fish were fed three meal sizes (close to satiation, 50% and 25%). GE was analyzed using a general power function, and data indicated that the square root function best described GE in brook trout independently of meal size. GER experiments performed at 12.5 °C, 13.2 °C, 15 °C, 16.8 °C, 18.6 °C and 20 °C revealed an exponential increased in GER from 12.5  $\degree$ C to 18.6  $\degree$ C and then a sharp decline at 20  $\degree$ C. The effect of dietary energy density was determined using meals of 12.6 kJ  $g^{-1}$ , 17.7 kJ  $g^{-1}$ , and 22  $kJ$   $g^{-1}$ . GER increased with decreasing energy density of the meal. Neither feed storage conditions nor the arrival of a second meal influenced GER. The GER model for brook trout fed commercial pellets was  $\frac{ds_t}{dt} = -0.000860L^{1.31}E^{-0.41}e^{0.08T}(1$  $e^{1.82(T-20.6)})\sqrt{S_t}$  (g h<sup>-1</sup>), where  $S_t$  is current stomach mass (g) and *t* is time (h). This GER model was used to calculate the amount of food that should be present in the stomach of a brook trout that had been fed on three consecutive days. The predicted values were similar to the observed ( $r^2 = 0.987$ ). The GER model can be used to predict stomach content in fish that have been fed repeatedly and should have applications in the development of feeding routines for brook trout.

## **Key Words**: Dietary Energy Density, Feeding Regime, Fish Size, General Power Function, Meal Size, Multiple Meals, Optimum Temperature

#### Doktora Tezi

## **ÖZET**

## <span id="page-8-0"></span>FORMÜLE YEM İLE BESLENEN KAYNAK ALABALIĞINDA MİDE BOŞALTIMI

#### Umar KHAN

Karadeniz Teknik Üniversitesi Fen Bilimleri Enstitüsü Balıkçılık Teknolojisi Mühendisliği Anabilim Dalı Danışman: Prof. Dr. Kadir SEYHAN 2020, 42 Sayfa, 12 Sayfa Ek

Balıklarda mide boşaltım oranı (GER) çalışmaları ile yetiştiriciliği yapılan balıkların yem alımının durduğu ve tekrar başladığı andaki mide doluluğu hesaplanarak optimum besleme sıklığı belirlenebilir. Bu çalışmada, ticari pelet yemlerle beslenen kaynak alabalıklarında; yem miktarı, balık boyu, sıcaklık, yemin enerji miktarı, yemin saklama koşulları ve ilave yemleme gibi faktörlerin GER üzerine etkisi belirlenmiştir. Bu amaçla; küçük (15 cm) ve büyük (23 cm) balıklar üç farklı öğün büyüklüğü ile beslenmiştir. Elde edilen verilerle mide boşaltımı basit üstel ilişki ile belirlenmiştir. Ayrıca besin büyüklüğünden bağımsız olarak mide boşaltımını en iyi tanımlayan modelin karekök model olduğu belirlenmiştir. Sıcaklığın etkisinin belirlenmesi amacıyla 12,5 °C, 13,2 °C, 15,0 °C, 16,8 °C, 18,6 °C ve 20,0 °C'de denemeler yapılmış; 12,5 °C-18,6 °C arasında GER'de eksponansiyel bir artış, 20 °C'de ise keskin bir düşüş gözlemnlenmiştir. Kaolin ile seyreltilerek 12,6 kJ g<sup>-1</sup>, 17,7 kJ g<sup>-1</sup> ve 22,0 kJ g<sup>-1</sup> enerji miktarına sahip üç yem formüle edilmiş ve bunların GER üzerine olan etkileri belirlenmiştir. Yemdeki enerji miktarı azaldıkça mide boşaltımı artış göstermiştir. Bununla birlikte, GER üzerine depolama koşullarının ve ilave yemlemenin etkisinin olmadığı tespit edilmiştir. Ticari pelet yemle beslenen balıklarda GER modeli  $\frac{dS_t}{dt} = -0.000860L^{1.31}E^{-0.41}e^{0.08T}(1$  $e^{1.82(T-20.6)})\sqrt{S_t}$  (gr sa<sup>-1</sup>) olarak belirlenmiştir. Modelin test edilmesi için üç gün boyunca düzenli olarak beslenen balıkların mide doluluk oranları model ile hesaplanmıştır. Hesaplanan değerlerin gerçek değerlere çok yakın olduğu tespit edilmiştir. Bu model yetiştiricilik faaliyetlerinde beslenme rutinlerinin geliştirilmesinde geniş bir kullanım alanı bulabilecektir.

## **Anahtar Kelimeler**: Diyet Enerji Miktarı, Beslenme Rejimi, Balık Büyüklüğü, Genel Üstel Model, Öğün Büyüklüğü, Çoklu Öğün, Optimum Sıcaklık

# **LIST OF FIGURES**

# **Page No**



Figure 11. The stomach content mass *S<sup>t</sup>* [of formulated feed in brook trout \(](#page-39-1)*Salvelinus fontinalis*[\) as predicted by the square root function](#page-39-1)  $St = S0 -$ 120.000860L1.31E – 0.41e0.08  $T(1 - e1.82T - 20.6)$ t and plotted against [actual values. The fitted curve of the relationship is](#page-39-1)  $y = 0.982x + 2.592$  ( $r^2 =$ 0.987).  $S_0$  (g) is meal size,  $L$  [\(cm\) total fish length,](#page-39-1)  $E$  (kJ g<sup>-1</sup>) feed energy density, and *T* [\(°C\) temperature..](#page-39-1) 27



## **LIST OF TABLES**

# **Page No**

<span id="page-11-0"></span>

# **LIST OF ABBREVIATIONS**



## <span id="page-13-0"></span>**1. GENERAL INTRODUCTION**

Information about food intake of predators is needed for estimation of mortality rates of their prey species. Studies on gastric evacuation rate (GER) have been used to estimate rates of food intake in wild fish, which enable quantitative investigation of the trophic links between predators and their prey [\(Jobling, 1981;](#page-48-0) [Seyhan and Grove, 1998;](#page-49-0) [Andersen, 2001;](#page-45-0) [Mychek-Londer and Bunnell, 2013\)](#page-48-1). Studies on GER have also been used to develop optimal feeding regimes for farmed fish [\(Grove et al., 1978;](#page-47-0) [Talbot, 1985;](#page-50-0) [Booth et al., 2008\)](#page-46-0), such as by examination of the progress of gastric emptying and the level of stomach fullness at return of appetite [\(Lee et al., 2000;](#page-48-2) [Riche et al., 2004\)](#page-49-1).

Providing feed to farmed fish at times that coincide with peak appetite may lead to improved growth and feed utilization, and reduce levels of feed waste [\(Bolliet et al., 2001;](#page-46-1) [Dwyer et al., 2002;](#page-47-1) [Booth et al., 2008\)](#page-46-0). The effects of feeding regime on the production of farmed fish have been examined in many empirical studies [\(Andrews and Page, 1975;](#page-45-1) [Hogendoorn, 1981;](#page-47-2) [Wang et al., 1998;](#page-50-1) [Xie et al., 2011\)](#page-51-0). Several studies have revealed limits to fish growth with increasing feeding frequency [\(Jobling, 1983;](#page-48-3) [Tsevis et al., 1992;](#page-50-2) [Dwyer](#page-47-1)  [et al., 2002;](#page-47-1) [Tian et al., 2015\)](#page-50-3), and this may be an indication that there are limitations imposed by stomach capacity and fullness. If this is the case, the optimal level of stomach fullness at which fish should be fed could be estimated using GER models.

#### <span id="page-13-1"></span>**1.1. Effects of Potential Predictor Variables on GER**

#### <span id="page-13-2"></span>**1.1.1. Meal Size**

The impact of meal size on GER of fish is a much-disputed topic in the literature, and several authors reported different impacts of meal size on GER. Several studies have demonstrated a positive correlation between GER and meal size [\(Windell, 1967;](#page-51-1) [Bagge,](#page-45-2)  [1977;](#page-45-2) [dos Santos and Jobling, 1995;](#page-47-3) [Andersen, 1998\)](#page-45-3). In contrast, no effect of meal size on GER was observed by [Bromley \(1988\)](#page-46-2) for whiting (*Merlangius merlangus*) and cod (*Gadus morhua*). [Ruggerone \(1989\)](#page-49-2) reported a negative correlation between GER and meal size when meal size was expressed relative to fish body size.

#### <span id="page-14-0"></span>**1.1.2. Fish Size**

Compared to meal size, the effects of fish size on GER are less disputed, although the magnitude of the effect of fish body size on GER reported for different species have been found to vary considerably [\(Jobling, 1981;](#page-48-0) [Bromley, 1994;](#page-46-3) [Seyhan, 1994;](#page-49-3) [Andersen, 1999\)](#page-45-4). In contrast, other studies using the same meal size relative to fish body size, have not identified any effects of body size on GER, such as it was reported for brown trout (*Salmo trutta*) [\(Elliott, 1972\)](#page-47-4).

#### <span id="page-14-1"></span>**1.1.3. Dietary Energy Contents**

Several studies have reported an increase in GER when the energy content of the meal was reduced, and this is known as a compensatory response [\(Rozin and Mayer, 1961;](#page-49-4) [Grove](#page-47-0)  [et al., 1978;](#page-47-0) [Jobling, 1987\)](#page-48-4). However, the magnitude of such changes in GER have been reported to be substantially higher by Andersen (2001, 2012) as compared to the results of [Temming and Herrmann \(2003\).](#page-50-4)

#### <span id="page-14-2"></span>**1.1.4. Temperature**

<span id="page-14-3"></span>Temperature is known to be among the major variables affecting physiological process rates including GER in ectotherms. Generally, the relationship between temperature and GER has been described using an exponential. However, the GER of several fish species including brook trout [\(Sweka et al., 2004\)](#page-50-5) and Atlantic cod [\(Tyler, 1970;](#page-50-6) [Andersen, 2012\)](#page-45-5) has been found to drop sharply between the optimum temperature and the upper thermal tolerance limit.

#### **1.1.5. Prey Resistance to The Digestive Processes**

The impact of prey resistance (e.g., robust exoskeletons, energy density of prey) are scarcely investigated in GER experiments. Generally, fish prey are considered homogenous with regard to resistance (Andersen and Beyer, 2005a,b).whereas prey such as crustaceans with robust exoskeletons are known to have two evacuation stages including an initial phase

with slow decomposition of surface materials such as chitin, and a second phase known to have a significantly higher GER when more easily digestible materials are exposed [\(Berens](#page-46-4)  [and Murie, 2008;](#page-46-4) [Couturier et al., 2013;](#page-46-5) [Andersen et al., 2016\)](#page-45-6).

## <span id="page-15-0"></span>**1.1.6. Feeding Regime**

According to a number of studies, the arrival of new food into the stomach of a fish that still contains a previous meal may accelerate the GER of the gastric residuum and slow down that of the new meal [\(Fletcher et al., 1984;](#page-47-5) [Persson, 1984;](#page-49-5) [dos Santos and Jobling,](#page-47-6)  [1992\)](#page-47-6). The inference that there is an acceleration of GER of the gastric residuum and a slowing down of GER of the second meal arises as a result of using of mass-dependent models of GER, rather than mechanistically driven, surface-dependent models, such as the one proposed by Andersen and Beyer (2005a,b).

### <span id="page-15-1"></span>**1.2. Methodology and the Experimental Design**

The GER experiments are mostly performed on individually held fish acclimated to laboratory conditions where the fish actively has started feeding in the testing environment. Several authors have also performed GER experiments on groups of fish such as sprat (*Sprattus sprattus*), Atlantic herring (*Clupea harengus*), and sardines (*Sardinops sagax*) since it was not possible to maintain such fish species individually [\(cited in Bernreuther et](#page-46-6)  [al., 2009\)](#page-46-6). Furthermore, some GER studies on groups of fish have been performed to compare their results with those obtained from individually maintained fish [\(Bascinar et al.,](#page-46-7)  [2016;](#page-46-7) [Bascinar et al., 2017\)](#page-46-8).

The acclimated fish are then deprived food for a certain time (e.g., 72 h) prior to the GER experiment to let them empty their stomachs. They are then fed a known amount of food. Starvation of fish between one and six days did not affect GER in brown trout, whereas longer periods of starvation slowed down the GER [\(Elliott, 1972\)](#page-47-4).

Several methods have been established to retrieve the stomach contents from experimental fish at different postprandial time to estimate their GER. The most frequently used approach is the so-called serial slaughter method, where the fish are killed using an overdose of anesthesia or by a blow to the head) and their stomachs dissected to recover the

stomach contents [\(Bromley, 1994;](#page-46-3) [Andersen, 2012\)](#page-45-5). Also, stomach lavage on anaesthetized fish has been used to recover the stomach contents without killing the fish [\(Seyhan et al.,](#page-49-6)  [1998;](#page-49-6) [Sweka et al., 2004\)](#page-50-5). Another method for studying GER, and especially total gastric emptying time (GET) is using X-radiography technique whereby experimental fish are fed formulated diets or natural prey that were incorporated with radio-opaque marker such as barium sulphate (BaSO4) powder or its suspension injected into prey body [\(Seyhan, 1994;](#page-49-3) [Mazlum and Alabdullah, 2019\)](#page-48-5). The movement of such feeds through the gut are then monitored by sequential X-ray photo of the gut at different postprandial times. However, the validity of radiographic methods depends on the markers passing through the gut of fish at a similar rate as the food, which is not always the case [\(Jørgensen and Jobling, 1988\)](#page-48-6). Actually, GET rather than GER is monitored, and GER is inferred by the relationship between GET and meal size. Another issue with this method is the stress of fish that might slow down GER in the experimental fish [\(Talbot, 1985\)](#page-50-0).

## <span id="page-16-0"></span>**1.3. General Information About Brook Trout**

Brook trout (*Salvelinus fontinalis*) is a commercially important fish species belonging to the family Salmonidae (Figure 1). As the genus name *Salvelinus* indicates, it is more closely related to chars such as dolly varden trout (*S. malma*) and arctic char (*S. alpinus*) than to species of the genus *Salmo*. Hence, it is also known as brook char [\(Ryther, 1997\)](#page-49-7).



The species name *fontinalis* from the Latin means "living in springs" since brook trout prefers cold, well-oxygenated, clean water and is sensitive to high water temperature and acidity [\(Warren et al., 2017\)](#page-50-7). Hence, the migration of brook trout within tributaries is strongly regulated by the ambient water temperature and they tend to migrate to the nearest

cold-water refugia when the water temperature goes above 18° C [\(Petty et al., 2012\)](#page-49-8). The upper lethal temperature limit reported for brook trout is  $24^{\circ}$  C to  $25^{\circ}$  C (Taniguchi et al., [1998;](#page-50-8) [Wehrly et al., 2007\)](#page-51-2).

Brook trout have been raised throughout Europe, Asia, Africa and South America by tribal, private, state and federal fish hatcheries for stocking purposes and fish markets [\(Fischer et al., 2009\)](#page-47-7). This fish species was brought to Turkey from Europe in the 1990s and have been raised in rainbow trout (*Oncorhynchus mykiss*) farms [\(Başçinar et al., 2003;](#page-46-9) [Innal](#page-48-7)  [and Erk'akan, 2006\)](#page-48-7). The growth rate of brook trout is slower than rainbow trout, which negatively affects the viability of brook trout markets [\(Fischer et al., 2009\)](#page-47-7). The commercial aquaculture market prefers fish with robust growth rate reaching market size in less time potentially increasing fish sale to the market [\(Fischer et al., 2009\)](#page-47-7). The commercial viability of brook trout farming can be increased if the fish achieve a growth rate like rainbow trout acquiring a similar size as rainbow trout within the same time period [\(Fischer et al., 2009\)](#page-47-7). This would eventually be advantageous for the fish-farmer and could expand the market share of brook trout. Consequently, further studies on the physiology and rearing techniques such as feeding regimes are needed to optimize the growth rate of brook trout.

## <span id="page-17-0"></span>**1.4. Aims and Objectives**

The major objectives of the work described in this thesis were to develop a GER model for brook trout subjected to experimental treatments involving differences in meal size, feeding regime, feed energy, fish size and water temperature. As such, the thesis incorporates studies that cover:

- 1. Modelling of the influence of meal size on the course of gastric evacuation with the aim of describing GER independently of meal size.
- 2. The influence of body size on GER
- 3. The influence of temperature on GER, with the aim of estimating the optimum temperature as well as the upper temperature limit for GER.
- 4. How GER is influenced by the energy density of the feed.
- 5. The effect of feed storage (frozen or at ambient temperature) on GER.
- 6. The GER of fish given single- and double-meals to examine the influence of the arrival of a second meal on GER of total stomach contents.

The results presented in the thesis should serve as a contribution to the development of improved feeding regimes for farmed brook trout. Additionally, the results may also provide a basis for the estimation of rates of food consumption of wild brook trout.

<span id="page-18-1"></span><span id="page-18-0"></span>

Figure 1. Brook trout (*Salvelinus fontinalis* Mitchill, 1814; Pisces, Salmonidae)

## **2. MATERIALS AND METHODS**

#### <span id="page-19-0"></span>**2.1. Fish Acquisition**

Farm raised brook trout of various body sizes were obtained from Prof. Dr. İbrahim OKUMUŞ Aquaculture Research & Production Unit at KTÜ Sürmene Faculty of Marine Sciences, Turkey. The fish were categorized in three groups according to their body size (small, medium and large). Each group was stocked in a separate holding tank (1200 l), and were kept under natural photoperiod. Fish had a continuous supply of fresh water and air bubbling to ensure the dissolved oxygen levels close to 100%. The fish in holding tanks were fed twice daily with commercial pellets (trout diets) acquired from Skretting Aquaculture (Table 1).

<span id="page-19-2"></span>Table 1. Composition and energy density of the commercial pellets used for farmed brook trout (*Salvelinus fontinalis*). Diet-II was diluted by 20% and 40% kaolin for experiment 8 and 9, respectively

	Used in	Nutrients value (crude, %)			$*Energy$	
Feed batch	experiments	Protein Fat Fibre Ash				$(kJ g^{-1})$
Diet-I	$1-6+10-18$ 44.0			21.0 3.9 9.0		22.10
$Diet-II$	$7 - 9$	37.O		23.8 2.5	63	22.32

\*determined using bomb calorimetry

## <span id="page-19-1"></span>**2.2. Acclimation Period to Experimental Conditions**

Brook trout of required body size were transferred to the laboratory where each fish was held separately in individual tanks (~100 l) for the GER trials (Figure 2, Table 2). The fish were fed once a day for 10–15 days to allow acclimation to the experimental conditions. The experimental tanks were cleaned once daily by siphoning. After 10–15 days of acclimation, brook trout that feed regularly under the laboratory conditions were selected for the GER experiments.



Figure 2. Individual tanks used for stocking brook trout (*Salvelinus fontinalis*) for gastric evacuation experiments

## <span id="page-20-1"></span><span id="page-20-0"></span>**2.3. Measuring of Satiation Meal Sizes and Time to Satiation**

The acclimated brook trout were starved for 72 h to allow full evacuation of their stomachs. They were then fed individually from a pre-weighted initial meal sizes to apparent station when they stopped taking further food on their own. The uneaten pellets were retrieved by siphoning and were dried at 60 °C using Ecocell Drying Oven [\(Figure 3\)](#page-21-1). Dry mass of uneaten pellets was subtracted from pre-weighted initial meal sizes to determine the

satiation meal size for each fish. The time from the beginning of feeding to deliberate cessation of feeding was considered as satiation time [\(Brett, 1971\)](#page-46-10).



Figure 3. Uneaten pellets collected from individual tanks were dried in an Ecocell Drying Oven

## <span id="page-21-1"></span><span id="page-21-0"></span>**2.4. Gastric Evacuation Experiments**

Brook trout, pre-acclimated to laboratory conditions, were fed a single meal of commercial pellets on empty stomachs (starved for at least 72 h) in all GE experiments excluding experiment 8. Brook trout in experiment 8 were fed with double meals wherein we provided the second meal 12 hours after the first meal. In majority of GE experiments, each brook trout quickly consumed all the offered feed, and uneaten pellets left after c. 30 minutes was recovered and subtracted from the initial meal size following the procedure explained in [Khan et al. \(2016\)](#page-48-8) (Table 1).

Their stomach contents  $S_t$  (g) were then subsequently sampled by dissection of fish at predetermined postprandial times *t* until the first empty stomach observed [\(Figure 4\)](#page-22-1). Prior to dissection of fish, the sampled fish were euthanized using benzocaine (150-350 mg/L), and the weight and total length (to the nearest cm) of each fish were measured. The retrieved  $S_t$  were dried at 60 °C using the same procedure mentioned-above.



Figure 4. Brook trout (*Salvelinus fontinalis*) were dissected to retrieve the contents of their stomachs

# <span id="page-22-1"></span><span id="page-22-0"></span>**2.5. Effects of Meal and Fish Sizes**

<span id="page-22-2"></span>Experiments 1–3 on large fish and 4–6 on small fish were conducted to determine the patterns of GE in brook trout and to parameterize the effects of meal and body sizes on GER of brook trout. The fish were fed on meal sizes close to satiation meal in experiment 1 and 4, while 50% and 25% of this meal size were fed to brook trout in experiments 2, 5 and 3, 6, respectively (Table 2).



Table 2. Basic data (mean ± S.E.) on gastric evacuation experiments performed on brook trout (*Salvelinus fontinalis*) fed a single meal or

first meal fed at 0600 hours, b second meal at 1800 hours.

a

11

## <span id="page-24-0"></span>**2.6. Effects of Dietary Energy**

Kaolin as inert material was added to the previously crushed commercial pellets to obtain diets with different energy contents (Table 2). Three diets with different energy levels were prepared following the procedure explained in [Khan and Seyhan \(2019\).](#page-48-9) These diets were used in experiments 7-9 to parameterize the effects of energy levels on the GER of brook trout.

#### <span id="page-24-1"></span>**2.7. Effects of Temperature and Feed Storage Conditions**

Experiment 10-15 were performed at different water temperatures to parameterize the influence of temperature on GER of brook trout. The experiments were performed in ambient water temperature at different times of the year (Table 2). The effect of feed storage condition (stored either frozen or under ambient conditions) on GER of brook trout were examined in experiment 16 [\(Khan and Seyhan, 2019\)](#page-48-9).

#### <span id="page-24-2"></span>**2.8. GER of Single- and Double- Meals**

Brook trout in experiment 17 was fed a single meal while fed double meals in experiment 18 to compare the GER of single and double meals. Brook trout in experiment 18 was fed a second meal after 12 hours of the first meal [\(Khan and Seyhan, 2019\)](#page-48-9).

## <span id="page-24-3"></span>**2.9. Forecasting Capability of GER Model**

The GER model summarized from the GE data of experiments 1–18 was used to estimate the stomach fullness of brook trout at time *t*. Brook trout of different body sizes were fed for three consecutive days, and their stomach contents were recovered on third day after two hours of the last meal [\(for details Khan and Seyhan, 2019\)](#page-48-9).

#### <span id="page-25-0"></span>**2.10. Analysis of GE Data**

The pattern of food evacuation from the stomach of brook trout independently of meal size was determined using the general power model:

$$
\frac{\mathrm{d}S_t}{\mathrm{d}t} = -\rho S_t^{\alpha} \quad , \tag{1}
$$

where  $\rho$  is the rate parameter constant ( $g^{1-\alpha} h^{-1}$ ),  $\alpha$  is the power parameter describing the course of food evacuation. The pattern of food evacuation is linear if  $\alpha = 0$ , a square root model if  $\alpha = \frac{1}{2}$ , a so-called surface-area dependent model if  $\alpha = \frac{2}{3}$ , and an exponential function if  $\alpha = 1$ .

The nonlinear regression and the iterative Marquardt method (NLIN procedure) of SAS, 9.04.01 was used to estimate the parameter of the general power model. Equation (1) was integrated over time *t* from the ingestion of the meal  $(t = 0)$  to total evacuation of the meal:

$$
S_t = S_0 (1 - S_0^{(a-1)} \rho (1 - a) t)^{1/(1-a)} + \varepsilon \quad , \tag{2}
$$

where *ε* is an error term.

The rate parameter  $\rho$  was expanded to account for the effect of fish body size (i.e., total length) on the GER of brook trout. The relationship between rate parameter and fish size was described using a power function.

$$
\rho = \rho_{LT} L^{\lambda} \,, \tag{3}
$$

Experiments 1–6 were performed to describe the course of GE in brook trout together with the effects of meal and fish sizes. However, these experiments were completed at an unintended temperature range of 15.1 °C to 18.2 °C. Therefore, the relationship between GE and temperature at this range was described by an exponential function (Bromley, 1994, Seyhan, 1994) as:

$$
\rho = \rho_{LT} L^{\lambda} e^{\delta T} \tag{4}
$$

where  $\lambda$  and  $\delta$  are parameters to be estimated.

The estimated value of  $\delta$  from experiments 1–6 was close to  $\frac{1}{2}$  suggesting the square root model to adequately describe the GE of brook trout independently of meal size:

$$
\frac{\mathrm{d}s_t}{\mathrm{d}t} = -\rho \sqrt{S_t} \tag{5}
$$

The rate parameter constant  $\rho$  likewise to equation (3) and (4) was expanded to account to the effects of fish size and temperature:

$$
\frac{\mathrm{d}s_t}{\mathrm{d}t} = -\rho_{LT} L^{1.31} \sqrt{S_t} \tag{6a}
$$

$$
\frac{\mathrm{d}s_t}{\mathrm{d}t} = -\rho_{LT} L^{1.31} \mathrm{e}^{\delta T} \sqrt{S_t} \tag{6b}
$$

Equation (6) was integrated over time *t* from ingestion of meal  $(t = 0)$  to total emptying of the meal:

$$
0 \le t \le 2\sqrt{S_0} \left(\rho_{LT} L^{1.31} e^{\delta T}\right)^{-1} \tag{7}
$$

The data analyses for experiments 1–6 proved that square root model best described the course of GE in brook trout independently of the size of the meal.

For the effects of energy density, the rate parameter *ρ<sup>L</sup>* was expanded to account to the effects of dietary energy density  $E$  (kJ  $g^{-1}$ ) and temperature  $T$  ( $\degree$  C)

$$
\frac{\mathrm{d}S_t}{\mathrm{d}t} = -\rho_{LE} L^{1.31} E^{-\mu} \sqrt{S_t} \tag{8}
$$

This equation was applied to GE data from experiment 7–9 to quantify the effects of *E* on GER by estimation of the exponent  $\mu$ .

The rate parameter *ρLE* was further expanded by adding a simple exponential function to account for the effect of *T* within the lower temperature range 12.5–18.6<sup>∘</sup> C (experiments  $10-14$ :

$$
\frac{\mathrm{d}S_t}{\mathrm{d}t} = -\rho_{LET} L^{1.31} E^{-\mu} \mathrm{e}^{\delta T} \sqrt{S_t} \tag{9}
$$

The relationship between temperature and GER throughout the experimental temperature range 12.5–20.0<sup>∘</sup> C (experiments 10–14) was then described by a temperature optimum model following the method proposed by [Andersen \(2012\):](#page-45-5)

$$
\frac{\mathrm{d}S_t}{\mathrm{d}t} = -\rho_{LET} L^{1.31} E^{-\mu} e^{\delta_1 T} (1 - e^{\delta_2 (T - T_u)}) \sqrt{S_t} \tag{10}
$$

where  $\delta_1$  and  $\delta_2$  are temperature coefficients, and  $T_u$  is the upper temperature limit where GE is believed to zero. The optimum temperature  $T_{opt}$  was then calculated by:

$$
T_{\rm opt} = T_{\rm u} - \delta_2^{-1} \ln \left[ (\delta_1 + \delta_2) \delta_1^{-1} \right] \tag{11}
$$

The parameter values of equations 5-10 were estimated by nonlinear regression and the iterative Marquardt method (NLIN procedure, SAS, 9.04.01). These equations were integrated over time *t* from ingestion of the meal  $(t = 0)$  to total emptying of the meal:

$$
\sqrt{S_t} = \sqrt{S_0} - \frac{1}{2}\rho t; \quad 0 \le t \le 2\sqrt{S_0} \rho^{-1},
$$
\n(12)

with the rate parameter  $\rho$  expanded according to equations 5–10.

The rate parameter  $\rho_L$  for experiment 17 and 18 was determined using equation (6b). For experiment 18, *t* in equation (1) was replaced by  $(t - \tau)$  to provide the displacement in time between ingestion of the first and second meal where  $\tau = 12$  h. Furthermore, the remains of the first meal at time *τ* (immediately after ingestion of the second meal) was estimated for the individual fish using the rate parameter value obtained from the single-meal experiment.

The one-way ANOVA was used to check for statistical differences among rate parameter values obtained from experiment 1–6, and 7–8. Furthermore, the *t*-test was used to test statistical differences between the rate parameter values obtained from experiments 15 and 16. The same procedure was applied to compare the GER of single-and double-meals experiments (experiment 17 and 18) to examine the influence of the arrival of a second meal on GE.



#### <span id="page-29-0"></span>**3. RESULTS**

#### <span id="page-29-1"></span>**3.1. Satiation Meal Size**

The mean satiation meal size estimated for large and small fish were 2% and 1.8% of their body weight. The preferred method of feeding for brook trout was to feed off the bottom, but it can also feed actively in the water column and from the surface [\(Table](#page-29-4) 3).

<span id="page-29-4"></span>Table 3. Values (mean  $\pm$  s.D.) of the major variables from experiments on brook trout (*Salvelinus fontinalis*) fed satiation meals of commercial pellets at 15,1 °C

Length $(cm)$	Body mass $(g)$		Satiation meal $(g)$ Satiation time (min)	Obs. $(n)$
$23.7 \pm 0.6$	$162.1 \pm 13.4$	$3.46 \pm 1.05$	$11.9 \pm 2.5$	20
$15.4 \pm 0.6$	$39.4 \pm 5.5$	$0.71 \pm 0.20$	$6.1 \pm 2.1$	20

## <span id="page-29-2"></span>**3.2. GE Patterns**

The use of general power model to combine GE data obtained with experiments 1-6 estimated 0.52 of the power parameter  $\alpha$  suggested that the square root model best describe the GE in brook trout [\(Table](#page-30-0) 4). The estimated value of  $\lambda$  and  $\delta$  were then fixed to estimate the *α* from experiment 1-3 and 4-6. Their estimated values of *α* were also closed to ½ that evinced the consistency of the results.

#### <span id="page-29-3"></span>**3.3. Effects of Temperature, Meal and Body Sizes**

Using the square root function, the values of length exponent  $\lambda$  and temperature coefficient  $\delta$  were estimated from combined data (experiment 1-6). Fixing  $\lambda$  and  $\delta$  at the estimated values obtained from combine GE data, the rate parameter  $\rho_{LT}$  were determined for experiments 1-6, 1-3, 4-6 and for each individual experiment ([Table](#page-30-1) 5). Their estimated values of  $\rho_{LT}$  ranged from 4.42 to 4.80 and did not differ significantly (ANOVA;  $F_{5, 84}$  = 0.485;  $P > 0.05$ ). Thus, the course of GE in brook trout fed with commercial pellets can be

adequately described by the square root model independently of meal size. The GE curves in [Figure](#page-31-1) 5 and 6 were determined using the square root model  $\sqrt{S_t} = \sqrt{S_0} - \frac{1}{2}$  $rac{1}{2}\rho t$ .

<span id="page-30-0"></span>Table 4. Parameter estimates  $\pm$  95% CI using the general power function  $\frac{ds_t}{dt}$  =  $-\rho_{LT}L^{\lambda}e^{\delta T}S_t^{\alpha}$  to combined data on gastric evacuation of meals of commercial pellets fed to brook trout (*Salvelinus fontinalis*)

Experiment number	$\alpha$	$\delta$	$\lambda$	$\rho_{LT}$ (×10 <sup>-4</sup> )	Adj. $r^2$	Obs.
1-6 combined (all fish)	$0.52\pm0.08$	$0.029 \pm 0.032$	$1.23 \pm 0.44$	$8.45 \pm 11.95$ 0.977		90
1-3 combined (large fish) $0.52 \pm 0.10$		$0.029$ (fixed)	$1.23$ (fixed)	$8.56 \pm 0.61$ 0.969		46
4-6 combined (small fish) $0.59 \pm 0.13$		$0.029$ (fixed)	$1.23$ (fixed)	$9.48 \pm 1.39$ 0.927		44

<sup>\*</sup>*L*, total fish length (cm); *T*, temperature ( $^{\circ}$ C); *S<sub>t</sub>*, stomach content mass (g) at postprandial time *t* (h); α,  $\delta$ ,  $\lambda$  and  $\rho$ *LT*, estimated model parameters

<span id="page-30-1"></span>Table 5. Parameter estimates  $\pm$  95% CI using the square root function  $\frac{ds_t}{dt}$  =

	$-\rho_{LT}L^{\lambda}e^{\delta T}\sqrt{S_t}$ to combined data on gastric evacuation of meals of commercial		
pellets fed to brook trout (Salvelinus fontinalis)			

Experiment number	δ	$\Lambda$	$\rho_{LT}$ (×10 <sup>-4</sup> )	Adjusted $r^2$	Obs.
1-6 combined (all fish)	$0.052 \pm 0.025$	$1.31 \pm 0.17$	$4.61 \pm 0.32$	0.972	90
1-6 combined (all fish)	$0.052$ (fixed)	$1.31$ (fixed)	$4.64 \pm 0.13$	0.972	90
1-3 combined (large fish)	$0.052$ (fixed)	$1.31$ (fixed)	$4.64 \pm 0.18$	0.962	46
4-6 combined (small fish)	$0.052$ (fixed)	$1.31$ (fixed)	$4.62 \pm 0.23$	0.919	44
1	$0.052$ (fixed)	$1.31$ (fixed)	$4.70 \pm 0.22$	0.972	16
2	$0.052$ (fixed)	$1.31$ (fixed)	$4.51 \pm 0.46$	0.854	15
3	$0.052$ (fixed)	$1.31$ (fixed)	$4.65 \pm 0.44$	0.808	15
$\overline{4}$	$0.052$ (fixed)	$1.31$ (fixed)	$4.80 \pm 0.34$	0.918	13
5	$0.052$ (fixed)	$1.31$ (fixed)	$4.52 \pm 0.37$	0.858	15
6	$0.052$ (fixed)	$1.31$ (fixed)	$4.42 \pm 0.61$	0.551	16

<sup>\*</sup>*L*, total fish length (cm); *T*, temperature ( ${}^{\circ}$ C); *S<sub>t</sub>*, stomach content mass (g) at postprandial time *t* (h);  $\alpha$ ,  $\delta$ ,  $\lambda$  and  $\rho_{LT}$ , estimated model parameters

Since the effect of temperature on GER of brook trout were estimated from a narrow range of temperature (15.1 to 18.2° C), therefore, the temperature dependency of GER was again parameterized with data from GE experiment numbers 10-15.



<span id="page-31-1"></span>Figure 5. Gastric evacuation data and estimated curves using the square root function for large brook trout (*Salvelinus fontinalis*) fed meals of 4.0 g ( $\bullet$ ), 2.0 g ( $\circ$ ), and 0.8 g ( $\bullet$ )



<span id="page-31-2"></span>Figure 6. Gastric evacuation data and estimated curves using the square root function for small brook trout (*Salvelinus fontinalis*) fed meals of  $0.8 \text{ g}$  ( $\bullet$ ),  $0.4 \text{ g}$  ( $\circ$ ), and  $0.2 \text{ g}$  ( $\bullet$ )

## <span id="page-31-0"></span>**3.4. Effects of Dietary Energy**

The GER of brook trout has increased as the dietary energy density (kcal/g) of commercial pellets reduced by the addition of kaolin [\(Figure](#page-33-0) 7). The estimated value of the energy exponent  $\mu$  obtained from the combined data of experiments 7 – 9 was 0.41  $\pm$  0.16 (± 95% CI) (Table 6).

<span id="page-32-0"></span>Table 6. Estimates  $\pm$  95% CI of the energy density exponent  $\mu$  and the rate parameter  $\rho_{LE}$  using the square root function  $\frac{ds_t}{dt} = -\rho_{LE} L^{1.31} E^{-\mu} \sqrt{S_t}$  to data on gastric evacuation of brook trout (*Salvelinus fontinalis*)

Experiment number	$\rho_{LE}(\times 10^{-3})$	$\mu$	Adjusted $r^2$	Observation $(n)$
$7 - 9$	$2.97 \pm 1.39$	$0.41 \pm 0.16$	0.990	39
$7 - 9$	$2.94 \pm 0.11$	$0.41$ (fixed)	0.990	39
	$2.90 \pm 0.22$	$0.41$ (fixed)	0.986	14
-8	$2.98 \pm 0.18$	$0.41$ (fixed)	0.987	13
9	$2.92 \pm 0.21$	$0.41$ (fixed)	0.989	12

The rate parameters  $\rho_{LE}$  estimated for individual experiment by fixing  $\mu$  at 0.41 in equation (2) were not statistically different (one-way ANOVA;  $F_{4, 116} = 0.003$ ;  $P > 0.05$ ). Consequently, using by  $\rho_L = \rho_{LE} E^{-0.41}$  a curve line was provided to combined data that depicted a close relationship ( $r^2 = 0.997$ ) between the values  $\rho_{LE}$  estimated for individual experiment and dietary energy content. The GE curves to the individual experiments  $7 - 9$ were then provided by  $S_t = \sqrt{S_0} - \frac{1}{2}$  $\frac{1}{2}$  0.00297L<sup>1.31</sup>E<sup>-0.41</sup>t [Figure](#page-33-1) 8.



<span id="page-33-0"></span>Figure 7. The influence of dietary energy density *E* on gastric evacuation rate as represented by the rate parameter ρ*<sup>L</sup>* of brook trout (*Salvelinus fontinalis*). The three values of  $\rho_L$  (estimates  $\pm$  95% C.I.) are obtained from fish fed a formulated feed diluted with 0% ( $\Box$ ), 20% (■) or 40% (■) kaolin. The gastric evacuation curve is achieved using the relationship  $\rho_L = \rho_{LE} E^{-0.41}$  to all data



<span id="page-33-1"></span>Figure 8. Gastric evacuation in brook trout (*Salvelinus fontinalis*) fed formulated feeds diluted with 0%  $\circ$ , 20%  $\circ$ ) or 40%  $\bullet$ ) kaolin. The gastric evacuation curves are obtained by use of the square root function  $S_t = \sqrt{S_0}$  – 1  $\frac{1}{2}$  0.00297L<sup>1.31</sup>E<sup>-0.41</sup>t to evacuation data. *S*<sub>0</sub> (g) is the meal size, *L* (cm) total fish length, and  $E$  (kJ  $g^{-1}$ ) energy density of the meal

#### <span id="page-34-0"></span>**3.5. Effects of Temperature and Feed Storage Conditions**

The value of energy exponent  $\mu$  was fixed at 0.41 in equations (9) and (10) before the impact of temperature on GER of brook trout was determined. The GER of brook trout increased as the temperature is raised from a minimum at 12.5° C to a maximum at 18.6° C followed by a sharp declined at  $20.0^{\circ}$  C [\(Table](#page-37-0) 7). Hence, the relationship between temperature and GER of brook trout within the lower range  $12.5^{\circ}$  C –  $18.6^{\circ}$  C (experiment 10–14) was described by the exponential model [equation (9)], which estimated 0.063  $\pm$ 0.013 for  $\delta$ . This estimated value of  $\delta$  corresponds to a  $Q_{10}$  value of 1.9. Using the parameterized exponential model  $\rho_{LE} = \rho_{LET} e^{0.063T}$  a curve line was provided to describe the relationship between GER of brook trout and temperature within the lower temperature range ( $r^2$  = 0.999, [Figure 9\)](#page-35-0). In order to determine the relationship between GER of brook trout and temperature both at lower and higher temperature ranges, the temperature optimum model was applied which estimated a similar value of 0.063 for  $\delta_I$  without retrieving any value for  $\delta_2$ . The value of  $\delta_1$  was then fixed at 0.08 in accordance to [Andersen \(2012\)](#page-45-5) prior to running the temperature optimum model to estimate the values of  $\delta_2$  and  $T_u$  [\(Table](#page-37-0) 7).

The parameterized temperature optimum model  $\rho_{LE} = \rho_{LET} e^{0.08T} \left( 1 - e^{1.82(T - 20.6)} \right)$ was used to provide a curve line to the GE data of brook trout obtained at lower and higher temperature ranges [\(Figure](#page-36-0) 10). This parameterized model provided a good fit to GE data  $(r^2)$  $= 0.975$ ). Furthermore, this parameterized model showed that the GER of brook trout increased exponentially as the temperature is raised to an optimum value at 18.9° C, and then began to decrease rapidly at higher temperatures, reaching zero near to 21° C [\(Figure](#page-35-0)  [9\)](#page-35-0).



<span id="page-35-0"></span>Figure 9. The relationship between temperature *T* and gastric evacuation rate as represented by the rate parameter  $\rho_{LE}$  (estimates  $\pm$  95% CI) of brook trout (*Salvelinus fontinalis*) fed a formulated feed. The exponential curves are acquired using the relationships  $\rho_{LE}$  =  $\rho_{LET} e^{0.063T}$ (-----) and  $\rho_{LE} = \rho_{LET} e^{0.08T}$  (---) to all data except those obtained from the highest temperature, and the optimum by use of = 0.08 (1 − 1.82(−20.6) ) (–––) to all data



<span id="page-36-0"></span>Figure 10. Gastric evacuation in brook trout (*Salvelinus fontinalis*) fed formulated feed at different temperatures. The curves are acquired using the relationship  $S_t =$  $\sqrt{S_0} - \frac{1}{2}$  $\frac{1}{2}$ 0.000860L<sup>1.31</sup>E<sup>-0.41</sup>e<sup>0.08</sup><sup>T</sup>(1 – e<sup>1.82(T-20.6)</sup>)t, where *S*<sub>0</sub> (g) is the meal size, *L* (cm) total fish length, *E* (kJ  $g^{-1}$ ) feed energy density, and *T* (°C) temperature



Table 7. Parameter values (estimates ± 95% CI) obtained by use of equations (a), (b) and (c) to gastric evacuation data on brook trout (*Salvelinus fontinalis*) fed commercial pellets at different temperatures.

<span id="page-37-0"></span>L (cm), total fish length; T (°C), temperature; S<sub>t</sub> (g), stomach content mass at postprandial time t (h);  $\delta$ ,  $\delta$ <sub>1</sub>,  $\delta$ <sub>2</sub>,  $\rho$ <sub>LE</sub> and  $\rho$ <sub>LTE</sub>, estimated parameters.

The estimated rate parameter *ρLE* of brook trout fed on frozen commercial pellets (experiment 16) was found to be significantly similar to the rate parameter of experiment 15  $(t$ -test,  $P > 0.05$ ; Table 7). This did not result in any influence of feed storage conditions on the GER in brook trout.

#### <span id="page-38-0"></span>**3.6. GER of Single- and Double-Meals**

There was also no significant difference between the rate parameters *ρ<sup>L</sup>* estimated for experiment 17 (fed single meal) and experiment 18 (fed double meal) (*t*-test, P > 0.05). The total stomach content made up of two meals was therefore evacuated at the same GER as if it had come from a single meal [\(Table](#page-38-2) 8).

<span id="page-38-2"></span>Table 8. Estimates  $\pm$  95% CI of the rate parameter  $\rho_L$  using the square root function  $ds_t$  $\frac{d\Delta_t}{dt} = -\rho_L L^{1.31} \sqrt{S_t}$  to data from gastric evacuation experiments on a single meal or double meal.

Experiment number	$\rho_L \, (\times 10^{-3})$	Adj. $r^2$ Observation ( <i>n</i> )
11 Single meal	$1.28 + 0.11$ 0.827	
12 Double meals	$1.28 + 0.15$ 0.807	

*L*, total fish length (cm); *St*, stomach content mass (g) of either single meal or double meal (composed of the remains of two meals) at postprandial time *t* (h).

On the basis of the above results, the GER model for brook trout can be summarized by:

$$
\frac{dS_t}{dt} = -0.000860L^{1.31}E^{-0.41}e^{0.08T}(1 - e^{1.82(T - 20.6)})\sqrt{S_t}
$$
\n(13)

#### <span id="page-38-1"></span>**3.7. Forecasting Capability of GER Model**

The summarized GE model of brook trout (equation 13) was used to predict brook trout's stomach fullness that had been fed for three consecutive days. The values of stomach fullness predicted by GE model of brook trout were closely in line with the observed values [\(Figure](#page-39-1) 11).



<span id="page-39-1"></span>Figure 11. The stomach content mass  $S_t$  of formulated feed in brook trout (*Salvelinus fontinalis*) as predicted by the square root function  $S_t$  =  $\sqrt{S_0} - \frac{1}{2}$  $\frac{1}{2}$ 0.000860L<sup>1.31</sup>E<sup>-0.41</sup>e<sup>0.08</sup><sup>T</sup>(1-e<sup>1.82(T-20.6)</sup> ) and plotted against actual values. The fitted curve of the relationship is  $y = 0.982x + 2.592$  ( $r^2 = 0.987$ ). *S*<sub>0</sub> (g) is meal size, *L* (cm) total fish length,  $E$  (kJ  $g^{-1}$ ) feed energy density, and  $T$  ( $^{\circ}$ C) temperature.

## <span id="page-39-0"></span>**3.8. GER of Re-pelleted Feeds and Normal Pellets**

The rate parameter of brook trout fed crushed re-pelleted feeds (experiment 7) was  $0.00115 \left( \pm 0.00009 \right)$  which was significantly different from the rate parameter of brook trout  $(0.00095 \pm 0.00008)$  fed on normal pellets (*t*-test, P < 0.05). Though both types of pellets possessed the same moisture contents which did not differ significantly ( $t$ -test,  $P > 0.05$ ).

#### <span id="page-40-0"></span>**4. DISCUSSION**

#### <span id="page-40-1"></span>**4.1. GE Pattern**

The use of general power model to combine GE data from experiments 1-6 evinced that the square root model adequately described the course of GE in brook trout. Similar to the present study, turbot (*Scophthalmus maximus*) were fed meals of different sizes composed of formulated diets and X-radiography were used to determine GE time [\(Grove](#page-47-8)  [et al., 1985\)](#page-47-8). The power value estimated from the GE data of turbot was 0.55 suggesting the square root model to adequately describe the GE of turbot. Furthermore, several studies have proven the capability of the square root model to adequately describe the GE of different fish species such as pikeperch *Stizostedion lucioperca*, coho salmon *Oncorhynchus kisutch* and different gadoids which were fed a suite of natural prey species [\(cited in Khan et al.,](#page-48-8)  [2016\)](#page-48-8). The capability of this model to describe the GE in fish has been supported by a mechanistical cylinder model based upon simple surface consideration of prey that evacuates isometrically by continuous successive peeling off [\(Andersen and Beyer, 2005b,](#page-45-7) [2005a\)](#page-45-8). The cylinder model considers the prey as homogeneous in terms of resistance to the digestive process and biochemical composition throughout the body and formulated feeds are even more homogenous compared to natural prey. The cylinder model assumes that the meal is as a cylinder in the fish stomach where the radius of the cylinder gets reduced by successive peeling off without affecting the end of the cylinder (Andersen and Beyer, 2005a, b).

#### <span id="page-40-2"></span>**4.2. Effects of Body Size**

The length exponent estimated for brook trout in this study was 1.31 which is consistent with earlier studies that ranged 1.3-1.4 estimated from different fish species such as haddock (*Melanogrammus aeglefinus*), Atlantic cod, whiting and saithe (*Pollachius virens*) [\(cited in Khan et al., 2016\)](#page-48-8). Though several studies have also reported lower values of *λ* for Atlantic cod and pikeperch [\(Khan et al., 2016\)](#page-48-8). The possible reason behind the lower estimation of fish size impact on GER of Atlantic cod and for pikeperch might be the fact that the larger fish were fed with larger size prey fish that generally had higher energy density compared to smaller prey [\(Pedersen and Hislop, 2001\)](#page-49-9). Whereas, the smaller fish in these studies were fed with prey of smaller size. Hence, several studies including results from this

thesis showed that GER gets depressed when the energy density of meal increased [\(Jobling,](#page-48-4)  [1987\)](#page-48-4). Along with length exponent, body mass exponent was also estimated for a possible comparison with values on body mass exponent from literature. The estimated value of mass exponent for brook trout was  $0.40 \pm 0.05$  (95% C.I.) that is also in line with the findings of aforementioned authors. However, several other studies have obtained a lower estimate of mass exponent for turbot, Atlantic cod and pikeperch (*Stizostedion lucioperca*) [\(Khan et al.,](#page-48-8)  [2016\)](#page-48-8).

#### <span id="page-41-0"></span>**4.3. Effect of Energy Density**

The GER of brook trout were increased by decreasing the dietary energy density of commercial pellets through the dilution of kaolin. The GER was increased by 12.92% and 26.69% with 20% and 40% kaolin dilution respectively. Such an increase in GER is demonstrated as a compensatory response for decreases in the energy content of diet [\(cited](#page-48-9)  [in Khan and Seyhan, 2019\)](#page-48-9) and this process is known to drive by feedback loops from the upper intestine [\(Jobling, 1987\)](#page-48-4). The energy exponent  $\mu$  estimated from experiment 7-9 is 0.41 which is in line with the finding of [Temming and Herrmann \(2003\)](#page-50-4) reporting a value of  $\mu$  = 0.45 for cod. However these values of energy exponent  $\mu$  are substantially lower than [\(Andersen, 2001;](#page-45-0) [Andersen, 2012\)](#page-45-5) who reported a value of 0.85 for whiting, Atlantic cod and saithe fed different fish prey and crustacean species. The lower estimation for  $\mu$  from experiment 7-9 than [\(Andersen, 2001;](#page-45-0) [Andersen, 2012\)](#page-45-5) findings might be the types of food since commercial pellets disintegrate more rapidly in the fish stomach than natural prey which might cause fish to partly lose control on the process of evacuation and the intestine of fish becomes overloaded [\(cited in Khan and Seyhan, 2019\)](#page-48-9).

#### <span id="page-41-1"></span>**4.4. Effects of Temperature**

The application of temperature optimum model to describe the dependency of temperature on GER proved that the GER of brook trout increased with a raise in temperature up to around  $18.9^{\circ}$  C, and above this point the GER of brook trout tends to sharply decline until it stops near to 21 $^{\circ}$  C. The optimum temperature limits estimated for growth rate is 16 $^{\circ}$ C and 20.2° C for metabolic rate of brook trout and above these temperature limit the growth and metabolic rates of brook trout begin to sharply decline [\(cited in Khan and Seyhan, 2019\)](#page-48-9). The lethal temperature range reported for brook trout is  $24^{\circ}$  C to  $25^{\circ}$  C (Taniguchi et al., [1998;](#page-50-8) [Wehrly et al., 2007\)](#page-51-2). The migration of wild brook trout within tributaries is also strongly regulated by the temperature of ambient water and the fish tend to migrate to the nearest cold-water refugia as the water temperature went beyond 18° C [\(Petty et al., 2012\)](#page-49-8). Similar the findings of this study, an optimum temperature levels for the GER of Atlantic cod, Atlantic herring (*Clupea harengus*) and European sprat (*Sprattus sprattus*) have been reported [\(cited in Khan and Seyhan, 2019\)](#page-48-9). [Temming \(1995\)](#page-50-9) developed the first optimum temperature model for GER of fish to determine the temperature dependency of GER at low and high temperature. His model was latter modified by [Andersen \(2012\)](#page-45-5) to fix a limitation [\(see Khan and Seyhan, 2019\)](#page-48-9). The optimum temperature as well as upper temperature ranges estimated by [Temming \(1995\)'](#page-50-9)s and [Andersen \(2012\)'](#page-45-5)s models are identical.

Compared with the results ranging from  $0.078$  to  $0.083$  ( $Q_{10} = 2.18 - 2.29$ ) that have been obtained by several other studies, the estimated temperature coefficient  $\delta$  of 0.063 and corresponding to  $Q_{10} = 1.9$  was in the low end. However, some other studies also reported lower values of δ from 0.032 ( $Q_{10}$  = 1.38) to 0.068 ( $Q_{10}$  = 2.0) [\(see Khan and Seyhan, 2019\)](#page-48-9). The low value of  $\delta$  in the present study might be due to missing GE data at lower temperatures since [Seyhan \(1994\)](#page-49-3) obtained a significantly higher value of δ (*Q*<sup>10</sup> increased from 0.14 to 1.97) when including data at further low temperatures. Thus, following [\(Andersen, 2012\)](#page-45-5) the value of  $\delta_1$  was fixed at 0.08, which is in accordance with the version of temperature optimum function that best describes the effect of temperature on GER at low as well as high temperatures. [Bernreuther et al. \(2009\)](#page-46-6) also used  $\delta_1$  fixed at 0.077 prior to application of the temperature optimum function on GER data of *S. sprattus* at temperatures between 7.5 °C and 21.5 °C.

#### <span id="page-42-0"></span>**4.5. Effects of Feed Storage Conditions**

If not properly stored, formulated fish feeds rapidly deteriorate and develop molds [\(Robb et al., 2013\)](#page-49-10). It is recommended to store them under cool, dry and well-ventilated conditions, and to avoid direct sunlight [\(Cruz, 1996;](#page-47-9) [Craig et al., 2017\)](#page-47-10). The possible effects of freezing the feed on GER was examined for brook trout fed frozen (–15.0 °C) and nonfrozen commercial pellets. Meals of both treatments were evacuated at similar rates. No effect of storage of the formulated feed on the digestive rate could therefore be demonstrated.

Also, Andersen (2012) found that freshly killed and previously frozen  $(-20 °C)$  sandeel (*Ammodytes tobianus*) fed to Atlantic cod were evacuated at a similar rates. Further, feed storage conditions did not significantly influence food intake and growth performance in rainbow trout (*Oncorhynchus mykiss*) fed for 42 days with frozen feed (stored at 10° C, - 1.1° C and -15° C) or non-frozen (stored at room temperature, 20.8° C) [\(Khan et al., 2018\)](#page-48-10).

#### <span id="page-43-0"></span>**4.6. Effects of Second Meal on GER**

The total stomach content composed of the two meals was evacuated by brook trout at a rate similar to that of a single meal, which is consistent with previously reported studies on other predator species [\(Elliott, 1972;](#page-47-4) [El-Shamy, 1976;](#page-47-11) [Temming, 2002\)](#page-50-10). Therefore, GER of the total stomach content of brook trout fed pelleted feeds seems to be appropriately described independently of meal size and feeding regime by the parameterized square root function (Equation 13). Authors generally claim that a second meal in the stomach tends to speed up the evacuation of first meal and delay that of the second meal [\(see Khan and](#page-48-9)  [Seyhan, 2019\)](#page-48-9). This apparent acceleration of GER of first meal and deceleration of the second one seems to be an artefact resulting from the use of mass-dependent rather than surface-dependent models (see [Khan et al., 2016;](#page-48-8) [Khan and Seyhan, 2019\)](#page-48-9).

## <span id="page-44-0"></span>**5. CONCLUSION**

<span id="page-44-1"></span>An expanded fully-parameterized GE model (equation 13), with a basis in the simple square root function, can be used to estimate the stomach fullness at return of appetite in brook trout subjected to a range of environmental and feeding conditions setting (feeding regime, feed characteristics, fish size and temperature). The model may have application in the development of feeding regimes for farmed brook trout, thereby reducing feed waste and the risk of pollution to recipient waters of farm effluent.

## **6. REFERENCES**

- <span id="page-45-3"></span>Andersen, N. G., 1998. The effect of meal size on gastric evacuation in whiting, Journal of Fish Biology, 52,4, 743-755.
- <span id="page-45-4"></span>Andersen, N. G., 1999. The effects of predator size, temperature, and prey characteristics on gastric evacuation in whiting, Journal of Fish Biology, 54,2, 287-301.
- <span id="page-45-0"></span>Andersen, N. G., 2001. A gastric evacuation model for three predatory gadoids and implications of using pooled field data of stomach contents to estimate food rations, Journal of Fish Biology, 59,5, 1198-1217.
- <span id="page-45-5"></span>Andersen, N. G., 2012. Influences of potential predictor variables on gastric evacuation in Atlantic cod *Gadus morhua* feeding on fish prey: parameterization of a generic model, Journal of Fish Biology, 80,3, 595- 612.
- <span id="page-45-8"></span>Andersen, N. G. and Beyer, J. E., 2005a. Gastric evacuation of mixed stomach contents in predatory gadoids: an expanded application of the square root model to estimate food rations, Journal of Fish Biology, 67,5, 1413-1433.
- <span id="page-45-7"></span>Andersen, N. G. and Beyer, J. E., 2005b. Mechanistic modelling of gastric evacuation in predatory gadoids applying the square root model to describe surfacedependent evacuation, Journal of Fish Biology, 67,5, 1392-1412.
- <span id="page-45-6"></span>Andersen, N. G., Chabot, D. and Couturier, C. S., 2016. Modelling gastric evacuation in gadoids feeding on crustaceans, Journal of Fish Biology, 88,5, 1886- 1903.
- <span id="page-45-1"></span>Andrews, J. W. and Page, J. W., 1975. The Effects of Frequency of Feeding on Culture of Catfish, Transactions of the American Fisheries Society, 104,2, 317- 321.
- <span id="page-45-2"></span>Bagge, O., 1977. Meal size and digestion in cod (*Gadus morrhua* L.) and sea scorpion (*Myoxocephalus scorpius* L.), Meddelelser fra Danmarks Fiskeri-og Havundersoegelser (Denmark).
- <span id="page-46-8"></span>Bascinar, N., Bascinar, N., Khan, U. and Seyhan, K., 2017. Effects of meal and body sizes on gastric evacuation rate in brook trout *Salvelinus fontinalis* (Mitchill, 1814) fed commercial pellets in group feeding, Indian Journal of Fisheries, 64,3, 50-54.
- <span id="page-46-7"></span>Bascinar, N., Bascinar, N. S., Seyhan, K. and Khan, U., 2016. The Effect of Temperature on the Rate of Gastric Evacuation in Brook Trout *Salvelinus fontinalis* Fed on Commercial Pellets: Group-Feeding, Pakistan Journal of Zoology, 48,6.
- <span id="page-46-9"></span>Başçinar, N., Okumuş, I. and Serezli, R., 2003. The Development of Brook Trout (Salvelinus fontinalis Mitchill, 1814) embryos during the yolk sac period, Turkish Journal of Zoology, 27,3, 227-230.
- <span id="page-46-4"></span>Berens, E. and Murie, D., 2008. Differential digestion and evacuation rates of prey in a warm‐temperate grouper, gag *Mycteroperca microlepis* (Goode & Bean), Journal of Fish Biology, 72,6, 1406-1426.
- <span id="page-46-6"></span>Bernreuther, M., Temming, A. and Herrmann, J. P., 2009. Effect of temperature on the gastric evacuation in sprat *Sprattus sprattus*, Journal of Fish Biology, 75,7, 1525-1541.
- <span id="page-46-1"></span>Bolliet, V., Azzaydi, M. and Boujard, T., 2001. Effects of feeding time on feed intake and growth, Food intake in fish, 233-249.
- <span id="page-46-0"></span>Booth, M. A., Tucker, B. J., Allan, G. L. and Fielder, D. S., 2008. Effect of feeding regime and fish size on weight gain, feed intake and gastric evacuation in juvenile Australian snapper *Pagrus auratus*, Aquaculture, 282,1-4, 104- 110.
- <span id="page-46-10"></span>Brett, J. R., 1971. Satiation time, appetite, and maximum food intake of sockeye salmon (*Oncorhynchus nerka*), Journal of the Fisheries Research Board of Canada, 28, 409–415.
- <span id="page-46-2"></span>Bromley, P., 1988. Gastric digestion and evacuation in whiting, *Merlangius merlangus* (L.), Journal of Fish Biology, 33,2, 331-338.
- <span id="page-46-3"></span>Bromley, P. J., 1994. The role of gastric evacuation experiments in quantifying the feeding rates of predatory fish, Reviews in Fish Biology and Fisheries, 4,1, 36-66.
- <span id="page-46-5"></span>Couturier, C. S., Andersen, N. G., Audet, C. and Chabot, D., 2013. Prey exoskeletons influence the course of gastric evacuation in Atlantic cod *Gadus morhua*, Journal of Fish Biology, 82,3, 789-805.
- <span id="page-47-10"></span>Craig, S., Helfrich, L. A., Kuhn, D. and Schwarz, M. H., 2017. Understanding fish nutrition, feeds, and feeding, Virginia: College of Agriculture and Life Sciences, Virginia Polytechnic Institute and State University. pp. 420-256.
- <span id="page-47-9"></span>Cruz, P. S., 1996. Feed quality problems and management strategies, Proceeding of the national seminar-Workshop on Fish Nutrition and Feeds, pp. 64-73.
- <span id="page-47-6"></span>dos Santos, J. and Jobling, M., 1992. A model to describe gastric evacuation in cod (*Gadus morhua* L.) fed natural prey, ICES Journal of Marine Science, 49,2, 145-154.
- <span id="page-47-3"></span>dos Santos, J. and Jobling, M., 1995. Test of a food consumption model for the Atlantic cod, ICES Journal of Marine Science, 52,2, 209-219.
- <span id="page-47-1"></span>Dwyer, K. S., Brown, J. A., Parrish, C. and Lall, S. P., 2002. Feeding frequency affects food consumption, feeding pattern and growth of juvenile yellowtail flounder (*Limanda ferruginea*), Aquaculture, 213,1-4, 279-292.
- <span id="page-47-11"></span>El-Shamy, F. M., 1976. Analyses of Gastric Emptying in Bluegill (Lepomis macrochirus), Journal of the Fisheries Research Board of Canada, 33,7, 1630-1633.
- <span id="page-47-4"></span>Elliott, J. M., 1972. Rates of gastric evacuation in brown trout, *Salmo trutta* L, Freshwater Biology, 2,1, 1-18.
- <span id="page-47-7"></span>Fischer, G. J., Held, J., Hartleb, C. and Malison, J., 2009. Evaluation of brook trout production in a coldwater recycle aquaculture system, Aquacultural engineering, 41,2, 109-113.
- <span id="page-47-5"></span>Fletcher, D., Grove, D., Basimi, R. and Ghaddaf, A., 1984. Emptying rates of single and double meals of different food quality from the stomach of the dab, *Limanda limanda* (L.), Journal of fish biology, 25,4, 435-444.
- <span id="page-47-8"></span>Grove, D., Moctezuma, M., Flett, H., Foott, J., Watson, T. and Flowerdew, M., 1985. Gastric emptying and the return of appetite in juvenile turbot, *Scophthalmus maximus* L., fed on artificial diets, Journal of Fish Biology, 26,3, 339-354.
- <span id="page-47-0"></span>Grove, D. J., Loizides, L. G. and Nott, J., 1978. Satiation amount, frequency of feeding and gastric emptying rate in *Salmo gairdneri*, Journal of Fish Biology, 12,5, 507-516.
- <span id="page-47-2"></span>Hogendoorn, H., 1981. Controlled propagation of the African catfish, *Clarias lazera* (C. & V.), Aquaculture, 24, 123-131.
- <span id="page-48-7"></span>Innal, D. and Erk'akan, F., 2006. Effects of exotic and translocated fish species in the inland waters of Turkey, Reviews in Fish Biology and Fisheries, 16,1, 39- 50.
- <span id="page-48-0"></span>Jobling, M., 1981. Mathematical models of gastric emptying and the estimation of daily rates of food consumption for fish, Journal of Fish Biology, 19,3, 245-257.
- <span id="page-48-3"></span>Jobling, M., 1983. Effect of feeding frequency on food intake and growth of Arctic charr, *Salvelinus alpinus* L, Journal of Fish Biology, 23,2, 177-185.
- <span id="page-48-4"></span>Jobling, M., 1987. Influences of food particle size and dietary energy content on patterns of gastric evacuation in fish: test of a physiological model of gastric emptying, Journal of Fish Biology, 30,3, 299-314.
- <span id="page-48-6"></span>Jørgensen, E. and Jobling, M., 1988. Use of radiographic methods in feeding studies: a cautionary note, Journal of fish biology, 32,3, 487-488.
- <span id="page-48-9"></span>Khan, U. and Seyhan, K., 2019. Gastric evacuation rates in farmed brook trout subjected to a range of feeding conditions fed commercial pellets, Aquaculture, 734390.
- <span id="page-48-8"></span>Khan, U., Seyhan, K., Başçinar, N. and Başçinar, N., 2016. Satiation meal and the effects of meal and body sizes on gastric evacuation rate in brook trout *Salvelinus fontinalis* fed commercial pellets, Journal of fish biology, 89,2, 1227-1238.
- <span id="page-48-10"></span>Khan, U., Terzi, Y. and Seyhan, K., 8th International Conference "Water & Fish", 13- 15 June 2018, Belgrade, Serbia, Growth metrices of juvenile rainbow trout as influenced by feed storage conditions (frozen or non-frozen): 163-163.
- <span id="page-48-2"></span>Lee, S.-M., Hwang, U.-G. and Cho, S. H., 2000. Effects of feeding frequency and dietary moisture content on growth, body composition and gastric evacuation of juvenile Korean rockfish (*Sebastes schlegeli*), Aquaculture, 187,3-4, 399-409.
- <span id="page-48-5"></span>Mazlum, R. E. and Alabdullah, A., 2019. Gastric evacuation in endangered Russian sturgeon (*Acipenser gueldenstaedtii*): Effects of body and meal sizes, Journal of Applied Ichthyology, 36,1, 39-45.
- <span id="page-48-1"></span>Mychek-Londer, J. G. and Bunnell, D. B., 2013. Gastric evacuation rate, index of fullness, and daily ration of Lake Michigan slimy (*Cottus cognatus*) and

deepwater sculpin (*Myoxocephalus thompsonii*), Journal of Great Lakes Research, 39,2, 327-335.

- <span id="page-49-9"></span>Pedersen, J. and Hislop, J., 2001. Seasonal variations in the energy density of fishes in the North Sea, Journal of Fish Biology, 59,2, 380-389.
- <span id="page-49-5"></span>Persson, L., 1984. Food evacuation and models for multiple meals in fishes, Environmental biology of fishes, 10,4, 305-309.
- <span id="page-49-8"></span>Petty, J. T., Hansbarger, J. L., Huntsman, B. M. and Mazik, P. M., 2012. Brook trout movement in response to temperature, flow, and thermal refugia within a complex Appalachian riverscape, Transactions of the American Fisheries Society, 141,4, 1060-1073.
- <span id="page-49-1"></span>Riche, M., Haley, D. I., Oetker, M., Garbrecht, S. and Garling, D. L., 2004. Effect of feeding frequency on gastric evacuation and the return of appetite in tilapia *Oreochromis niloticus* (L.), Aquaculture, 234,1-4, 657-673.
- <span id="page-49-10"></span>Robb, D. H., Crampton, V. O., Robb, D. and Crampton, V., 2013. On-farm feeding and feed management: perspectives from the fish feed industry, On-farm feeding and feed management in aquaculture, 489-518.
- <span id="page-49-4"></span>Rozin, P. and Mayer, J., 1961. Regulation of food intake in the goldfish, American Journal of Physiology, 201,5, 968-974.
- <span id="page-49-2"></span>Ruggerone, G., 1989. Gastric evacuation rates and daily ration of piscivorous coho salmon, *Oncorhynchus kisutch* Walbaum, Journal of Fish Biology, 34,3, 451-463.
- <span id="page-49-7"></span>Ryther, J. H., 1997. Anadromous brook trout: biology, status and enhancement, Wood Hole Oceanographic Institution, MA, Trout Unlimited Report.
- <span id="page-49-3"></span>Seyhan, K., 1994. Gastric emptying, food consumption and ecological impact of whiting, *Merlangius merlangus* (L.) in the eastern Irish Sea marine ecosystem, University of Wales, Bangor, United Kingdom. pp. 59-60.
- <span id="page-49-6"></span>Seyhan, K., Başçinar, N. S. and DÜZGÜNEŞ, E., 1998. Some Observations on Gastric Emptying in the Black Sea Whiting, *Merlangius merlangus euxinus*, (N. 1840) Fed on Natural Prey, Turkish Journal of Zoology, 22,3, 251-258.
- <span id="page-49-0"></span>Seyhan, K. and Grove, D. J., 1998. Food consumption of whiting, *Merlangius merlangus*, in the Eastern Irish Sea, Fisheries Research, 38,3, 233-245.
- <span id="page-50-5"></span>Sweka, J. A., Keith Cox, M. and Hartman, K. J., 2004. Gastric evacuation rates of brook trout, Transactions of the American Fisheries Society, 133,1, 204- 210.
- <span id="page-50-0"></span>Talbot, C., 1985. Fish energetics, Laboratory methods in fish feeding and nutritional studies, Springer, 125-154.
- <span id="page-50-8"></span>Taniguchi, Y., Rahel, F. J., Novinger, D. C. and Gerow, K. G., 1998. Temperature mediation of competitive interactions among three fish species that replace each other along longitudinal stream gradients, Canadian Journal of Fisheries and Aquatic Sciences, 55,8, 1894-1901.
- <span id="page-50-9"></span>Temming, A., 1995. Die quantitative Bestimmung der Konsumtion von Fischen. Experimentelle, methodische und theoretische Aspekte (in German), Universität Hamburg Habilitationsschrift.
- <span id="page-50-10"></span>Temming, A., 2002. Estimating the mean time between meals in the field from stomach content data and gastric evacuation functions of whiting (*Merlangius merlangus* L.) feeding on sandeel (*Ammodytes marinus Raitt*), ICES Journal of Marine Science, 59,4, 782-793.
- <span id="page-50-4"></span>Temming, A. and Herrmann, J. P., 2003. Gastric evacuation in cod, Fisheries Research, 63,1, 21-41.
- <span id="page-50-3"></span>Tian, H.-Y., Zhang, D.-D., Li, X.-F., Zhang, C.-N., Qian, Y. and Liu, W.-B., 2015. Optimum feeding frequency of juvenile blunt snout bream *Megalobrama amblycephala*, Aquaculture, 437, 60-66.
- <span id="page-50-2"></span>Tsevis, N., Klaoudatos, S. and Conides, A., 1992. Food conversion budget in sea bass, *Dicentrarchus labrax*, fingerlings under two different feeding frequency patterns, Aquaculture, 101,3-4, 293-304.
- <span id="page-50-6"></span>Tyler, A. V., 1970. Rates of Gastric Emptying in Young Cod, Journal of the Fisheries Research Board of Canada, 27,7, 1177-1189.
- <span id="page-50-1"></span>Wang, N., Hayward, R. S. and Noltie, D. B., 1998. Effect of feeding frequency on food consumption, growth, size variation, and feeding pattern of age-0 hybrid sunfish, Aquaculture, 165,3-4, 261-267.
- <span id="page-50-7"></span>Warren, D. R., Kraft, C. E., Josephson, D. C. and Driscoll, C. T., 2017. Acid rain recovery may help to mitigate the impacts of climate change on thermally sensitive fish in lakes across eastern North America, Global change biology, 23,6, 2149-2153.
- <span id="page-51-2"></span>Wehrly, K. E., Wang, L. and Mitro, M., 2007. Field‐based estimates of thermal tolerance limits for trout: incorporating exposure time and temperature fluctuation, Transactions of the American Fisheries Society, 136,2, 365- 374.
- <span id="page-51-1"></span>Windell, J., Rates of digestion in fishes; in the biological basis of freshwater fish production ed. SD Gerking*.* pp. 151–173. Oxford: Blackwell Scientific. (1967).
- <span id="page-51-0"></span>Xie, F., Ai, Q., Mai, K., Xu, W. and Ma, H., 2011. The optimal feeding frequency of large yellow croaker (*Pseudosciaena crocea*, Richardson) larvae, Aquaculture, 311,1-4, 162-167.

## <span id="page-52-0"></span>**7. APPENDIX**

## <span id="page-52-1"></span>**7.1. General Power Model**

SAS program codes that were used to estimate the parameters of equation-4 (General power model):

FILENAME CSV "/folders/myfolders/PhD\_GE all experiments.csv" TERMSTR=CRLF;

## **PROC IMPORT** DATAFILE=CSV

 OUT=Brooktrout DBMS=CSV REPLACE;

**RUN**;

title 'Gastric evacuation in brook trout. +AC model'; **data** exp; set Brooktrout; if expno=**1** or expno=**2** or expno=**3** or expno=**4** or expno=**5** or expno=**6**; **run**;

**proc nlin** data=exp method=marquardt; parms A=**0.08** B=**0.5** C=**1.5** RAC=**0.0005**; delta=**0.00000001**;

R=RAC\*EXP(A\*temp)\*predlcm\*\*C;

s=sow\*\*(**1**-B)-R\*(**1**-B)\*time; if s>0 then stx=s\*\*( $1/(1-B)$ ); else stx=**0**; model stw=stx;

```
output out=expp p=pstw r=stw_residual;
run;
proc summary data=expp;
    var stw_residual stw;
   output out=stats css(stw)=sstot uss(stw_residual)=ssres N=N;
run;
data expp;
  set stats;
   rsquared=1-(ssres/sstot);
    adjrsquared = 1-(1-rsquared)*(N-1) / (N- 4 -1);
run;
proc print data=expp;
run;
```
Here "a" stands for *δ* (temperature), "b" for *α* (shape exponent) and "c" for *λ* (exponent of fish length), and RAC for rate parameter constant *ρLT*.

## <span id="page-53-0"></span>**7.2. Square Root Model**

SAS program codes that were used to estimate the parameters of equation 10:

FILENAME CSV "/folders/myfolders/PhD\_GE all experiments.csv" TERMSTR=CRLF;

## **PROC IMPORT** DATAFILE=CSV

 OUT=Brooktrout DBMS=CSV REPLACE;

**RUN**;

title 'Gastric evacuation in brook trout. Temperature effect+E'; **data** exp; set Brooktrout;

```
if expno=10 or expno=11 or expno=12 or expno=13 or expno=14 or expno=15;
run;
```
**proc nlin** data=exp method=marquardt;

parms RLTE=**0.0022** A=**0.08** B=**0.46** C=**1.31** E=**0.41** TU=**20**;

R=RLTE\*predlcm\*\*C\*EXP(A\*temp)\*(**1**-EXP(B\*(temp-TU)))\*enerd\*\*-E;

model sqrtstw=sqrtsow-**0.5**\*R\*time;

output out=expp p=psqrtstw r=stw\_residual;

**run**;

```
proc summary data=expp;
```

```
var stw residual stw;
```

```
 output out=stats css(stw)=sstot uss(stw_residual)=ssres N=N;
```
#### **run**;

```
data expp;
  set stats;
   rsquared=1-(ssres/sstot);
   adjrsquared = 1-(1-rsquared)*(N-1) / (N-3 -1);
run;
proc print data=expp;
```
## **run**;

Here "a" stands for  $\delta_I$  (1<sup>st</sup> temperature coefficient), "b" for  $\delta_2$  (2<sup>nd</sup> temperature coefficient), "c" for  $\lambda$  (exponent of fish length), E for  $\mu$  (coefficient of energy density),  $T_U$ upper temperature limit, and RLTE for rate parameter constant *ρLET*.

- <span id="page-55-0"></span>**7.3. Raw Data Obtained from the Gastric Evacuation Experiments Listed in Table 2.**
- <span id="page-55-1"></span>**7.3.1. Experiment 1-6 for Parameterizing the Effects of Body and Meal Sizes on GE of Brook Trout**









## <span id="page-58-0"></span>**7.3.2. Experiment 7-9 for Parameterizing the Effects of Dietary Energy on Gastric Evacuation Rates of Brook Trout.**



## <span id="page-59-0"></span>**7.3.3. Experiment 10-15 for Parameterizing the Effects of Temperature on GER of Brook Trout.**









## <span id="page-62-0"></span>**7.3.4. Experiment 16 for Parameterizing the Effects of Feed Storage Condition on GER of Brook Trout.**





# <span id="page-63-0"></span>**7.3.5. Experiment 17-18 for Comparing the GER of Single and Double Meals Fed to Brook Trout.**

#### **CURRICULUM VITAE**

Umar KHAN graduated from the Government Higher Secondary School No.1 Peshawar Cantt., Pakistan in 2005. He received his B.Sc. in Biological Sciences and M.Sc. in Zoology (Specialization: Fisheries) from the University of Peshawar, Pakistan in 2009 and 2011, respectively. Until 2013, he worked as a senior biology teacher in a higher school where he taught GCSE O & A level. In 2013, he received a fully funded Ph.D. Scholarship from Türkiye Bursları (government-funded, competitive scholarship program), awarded to outstanding students to pursue a degree full-time at Turkish universities. His research interests include aquaculture and quantification of trophic links between predators and prey. He is fluent in five languages: English, Turkish, Pashto, Persian, and Urdu/Hindi. His publications which are SCI indexed include:

From current PhD thesis:

- 1. **Khan, U.,** Seyhan, K. (2019). Gastric evacuation rates in farmed brook trout subjected to a range of feeding conditions fed commercial pellets. Aquaculture, 513, 734390.<http://bit.ly/2QL22pD>
- 2. **Khan, U.,** Seyhan, K., Başçinar, N. and Başçinar, N. S. (2016). Satiation meal and the effects of meal and body sizes on gastric evacuation rate in brook trout *Salvelinus fontinalis* fed commercial pellets. *Journal of Fish Biology*, 89, pp.1227–1238. <http://bit.ly/2CTtPwP>

Some other publications:

- 3. Seyhan, K., Başçinar, N. S., Başçinar, N. and **Khan, U.** (2020). Gastric Evacuation in Brook Trout (*Salvelinus fontinalis*) Fry: Effect of Body Size. *Pakistan Journal of Zoology*.<http://bit.ly/34xfUr6>
- 4. Alkan, N., Terzi, Y., **Khan, U.,** Bascinar, N., and Seyhan, K. (2019). Evaluation of Seasonal Variations in Surface Water Quality of the Caglayan, Firtina and Ikizdere Rivers from Rize, Turkey. Fresenius Environmental Bulletin, 28 (12A), pp. 9679- 9688.<http://bit.ly/2PKkoVC>
- 5. Başçinar, N. S., Başçinar, N., **Khan, U.** and Seyhan, K. (2017). The effects of meal and body sizes on gastric evacuation rate in brook trout *Salvelinus fontinalis* fed commercial pellets: group-feeding. *Indian Journal of Fisheries*, 64(3), pp. 50-54. <http://bit.ly/2AyFijh>
- 6. Başçinar, N., Başçinar, N. S., Seyhan, K. and **Khan, U.** (2016). The Effect of Temperature on the Rate of Gastric Evacuation in Brook Trout *Salvelinus fontinalis* Fed on Commercial Pellets: Group-Feeding. *Pakistan Journal of Zoology*, 48(6), pp. 1899-1904.<http://bit.ly/2r7c0nq>
- 7. Önder, M.Y. and **Khan, U.** (2016). Effects of Mono-and Duoculture on the Survival and Growth Rate of Juvenile Abant Trout (*Salmo trutta abanticus*) and Brook Trout (*Salvelinus fontinalis*). *Pakistan Journal of Zoology*, 48(3), pp.895-898. <http://bit.ly/2zqOndY>
- 8. Önder, M.Y., Başçınar, N., **Khan, U.** and Sonay, F.D. (2016). Effect of Photoperiod on Growth and Efficiency of Yolk-Sac Utilization in Alevins of Brook Trout (*Salvelinus fontinalis*). *Pakistan Journal of Zoology,* 48(2), pp.533-537. <http://bit.ly/2zrBMqV>
- 9. **Khan, U.,** Hasan, Z., Inayatullah, M., and Jan, A. (2014). Feeding habits of a freshwater catfish, *Clupisoma naziri* (Pisces: Schilbidae) from Khyber Pakhtunkhwa rivers, Pakistan. *Pakistan Journal of Zoology,* 46(4), 1166-1169. <http://bit.ly/2Fbjyhg>
- 10. **Khan, U.,** Hasan, Z., Inayatullah, M., and Jan, A. (2013). Analysis of stomach contents of freshwater catfish, *Eutropiichthys vacha* (Hamilton, 1822) from Khyber Pakhtunkhwa Rivers, Pakistan. *Pakistan Journal of Zoology, 45*(4), 1153-1156. http://bit.ly/2GXPg46